

# Advanced Imaging for Tuberculosis: Insights into Disease Pathogenesis

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**Unite to End TB by Making the Connections**  
**Maryland Department of Health and Mental Hygiene**  
**March 9, 2017**

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# Disclosures

- Grant funding: NIH  
Gilead Biosciences
- Patent: International patent PCT/US13/59897, 'Bacteria-specific labeled substrates as imaging biomarkers to diagnose, locate and monitor infections' filed

**Sanjay K Jain**

# Objectives

- Describe a clinical vignette to demonstrate the power of imaging in monitoring drug-resistant TB
- Discuss limitations of current tools for diagnosis and monitoring of infections, and how imaging could overcome some of these limitations
- Discuss “perceived” and real risks of clinically available imaging modalities
- Describe two molecular imaging studies at Hopkins currently enrolling TB patients, and approved by the Maryland Health Department
- Describe some bacteria-specific imaging agents currently in development

# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child

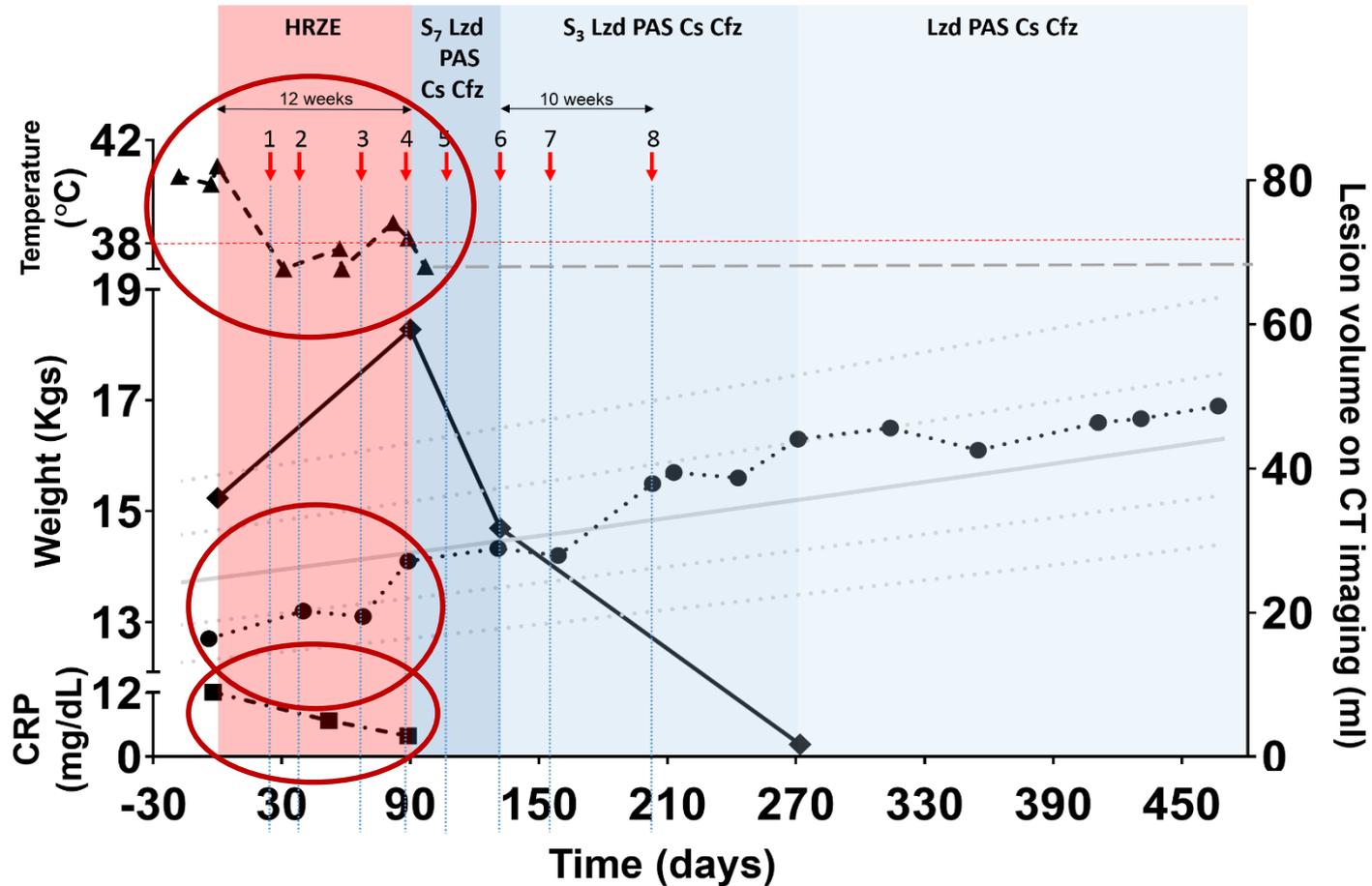
A two year-old child from the United States developed pneumonia after a three-month visit to India:

- On arrival in India, she was immunized with BCG, stayed with her grandparents and attended a local day-care facility. During the last week of her visit, she developed fevers that continued after her return to the USA.
- Based on clinical presentation and CT imaging findings, she was started on empiric first-line TB treatment after obtaining sequential gastric-aspirates.
- Symptoms resolved with first-line TB treatment but extensively drug-resistant (XDR) *Mycobacterium tuberculosis* grew in culture.
- The child was started on an individualized drug regimen for XDR-TB consisting of intravenous streptomycin, linezolid, PAS, cycloserine and clofazimine.
- But treatment was complicated by uncertainties about drug selection, and lack of child-friendly formulations. ***So, we needed to know (quickly), whether the individualized drug regimen was working or not!!!***
- However, assessing response to treatment was challenging, as clinical response was noted with suboptimal regimens, and microbiology could also not be used in this child.

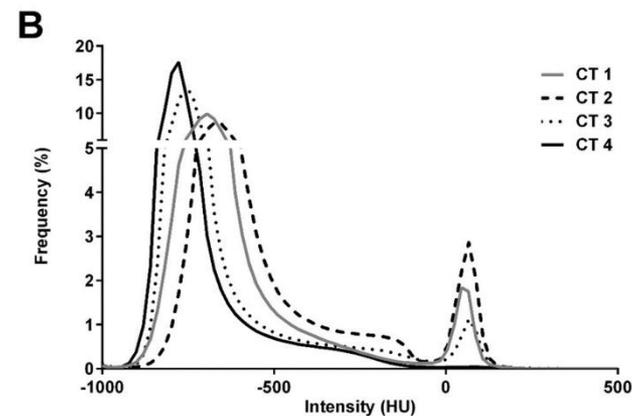
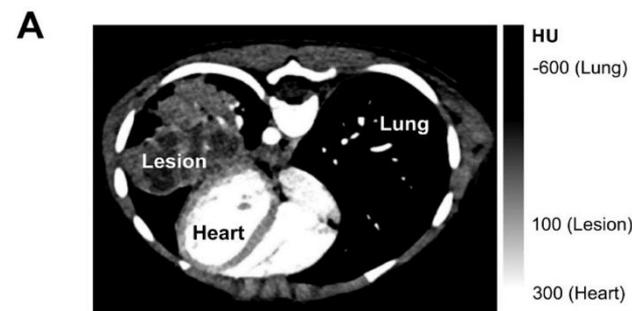
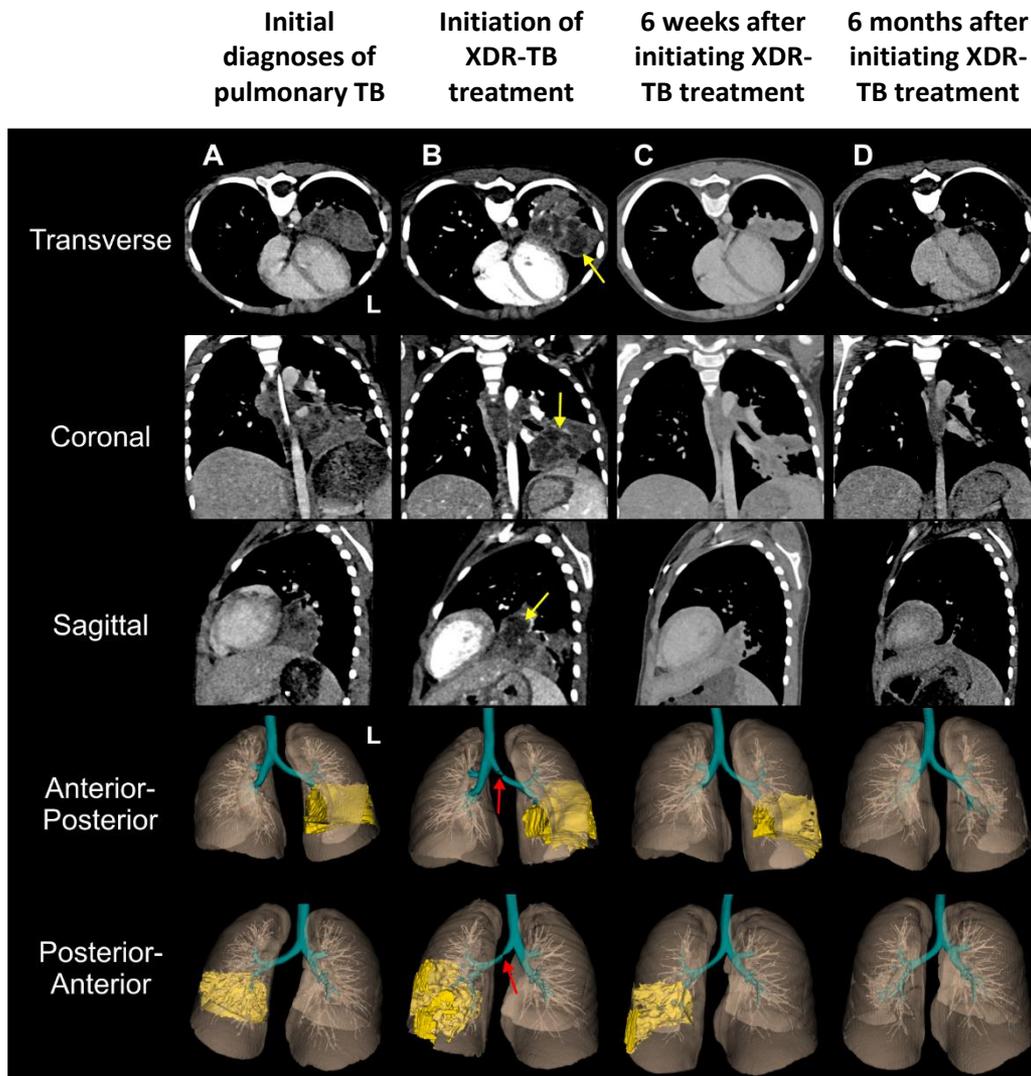
# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child

- Clinical response was noted with a suboptimal regimen.
- Gastric aspirate smears were negative at the time of initiation of XDR-TB treatment, and cultures remained negative subsequently.
- Therefore, in the absence of clinical or microbiological markers, low-radiation exposure pulmonary CT imaging was used to monitor treatment response, and guide the individualized drug regimen for XDR-TB.

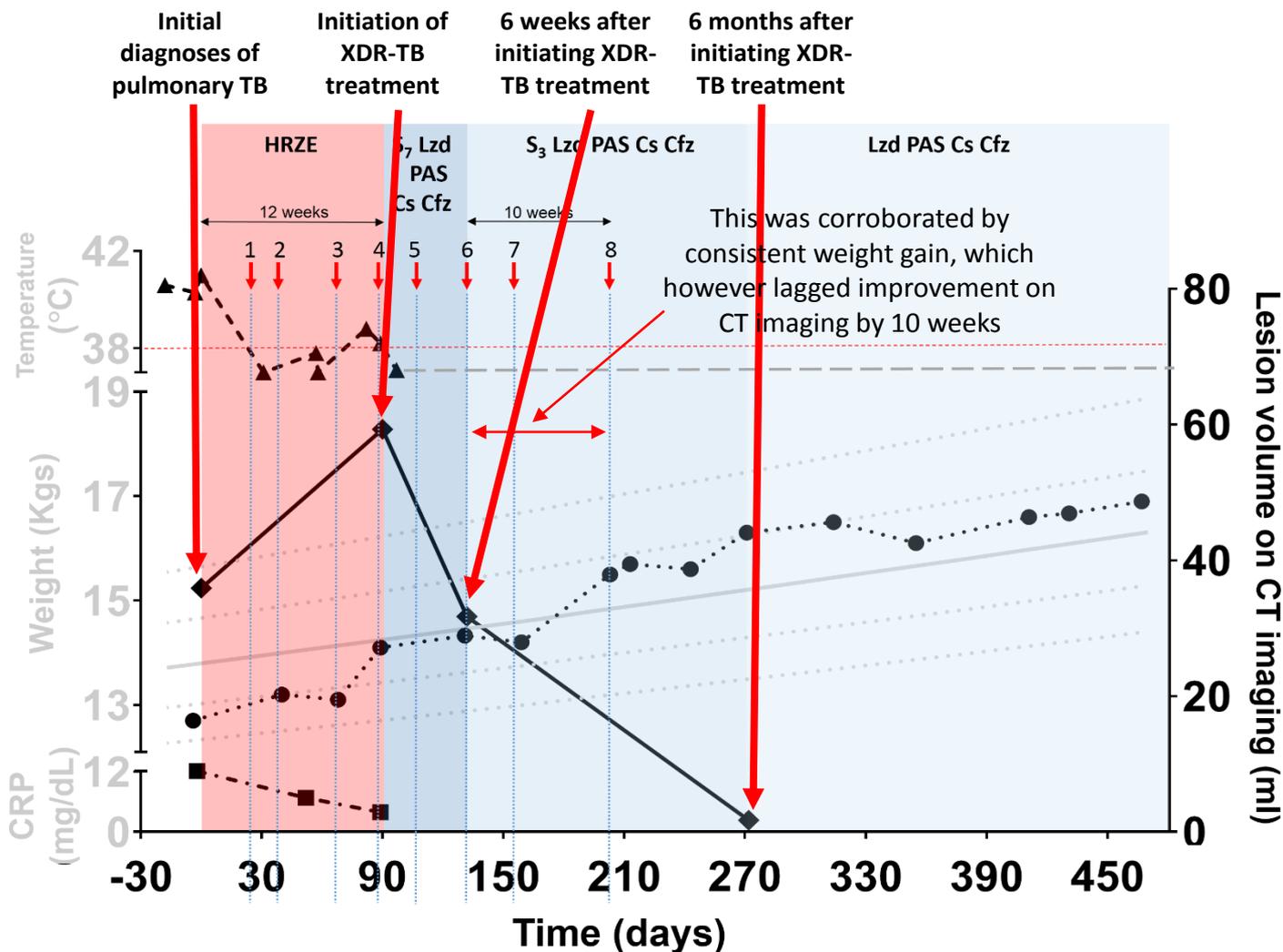
# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child



# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child



# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child



- 1, Growth of acid-fast bacilli in liquid broth;
- 2, Identification of *M. tuberculosis* complex by 16S rRNA sequencing;
- 3, Persistent left lower lobe infiltrate on chest radiography;
- 4, Sanger sequencing and initial TREK panel confirming XDR strain;
- 5, Agar proportion results (Maryland State);
- 6, CT demonstrating significant reduction in lesion volume;
- 7, Agar proportion results (National Jewish);
- 8, Consistent weight gain.

# CT Imaging to monitor Extensively drug-resistant (XDR) TB in a young child

- This report highlights the:
  - risks of acquiring drug-resistant (DR) TB overseas
  - *challenges of diagnosing DR-TB, especially in young children*
  - lack of dosing and child-friendly drug formulations
  - controversy regarding the infectious risk of young child and
  - *lack of tools to rapidly monitor DR-TB treatments*
- In this child, low-radiation exposure pulmonary CT demonstrated marked improvement as early as 6 weeks after initiation of treatment for XDR-TB, and corroborated by consistent weight gain, which however lagged improvement on CT imaging by 10 weeks.
- ***Treatments for drug resistant TB are often empiric, and the risks of treatment failure substantially outweigh the use of relatively advanced, but promising technologies that can provide rapid and reliable means to monitor treatments.***

# Food for thought . . .

- Most experts agree that Infections will not be eradicated for decades, or maybe never . . .
- Are we planning for the future, and utilizing the advances in technology that may be applicable to Infectious Diseases?
- Does diagnosis and monitoring of infections in special situations (hard to get locations, difficult to grow bugs, e.g. *M. tuberculosis* or other fastidious bugs) merit the development and / or use of technologies, that may be different from those being developed currently?

# The future is not what it used to be . . .



**Fundamental diagnostic: 1882**

**Need to isolate the bug**



**Fundamental diagnostic: 2017**

**Need to isolate the bug**



**MGIT**

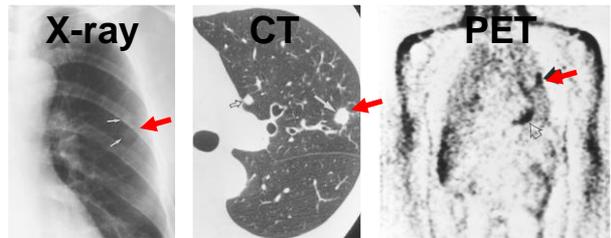
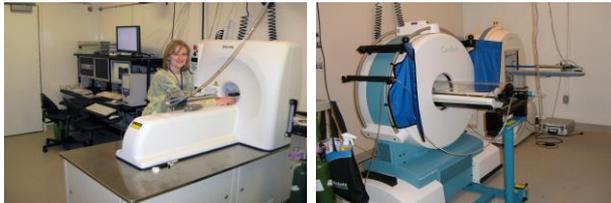
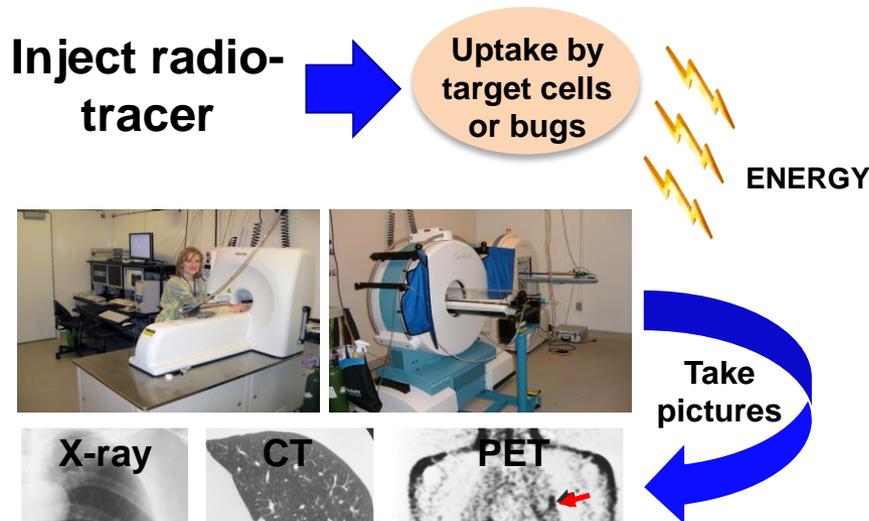


**GeneXpert**

# Problem(s) – Counting bugs

- Bugs: “need to bring them out, or go in and get them”
- Often impractical or dangerous
- Whole organ / body view of disease not available

# Solution(s) – Take a picture instead



<http://www.musc.edu/fnrd/primary.psych.images/page%2053.GIF>  
Goo JM et al. Radiology 2000



- Rapid, and *easily scalable to humans*

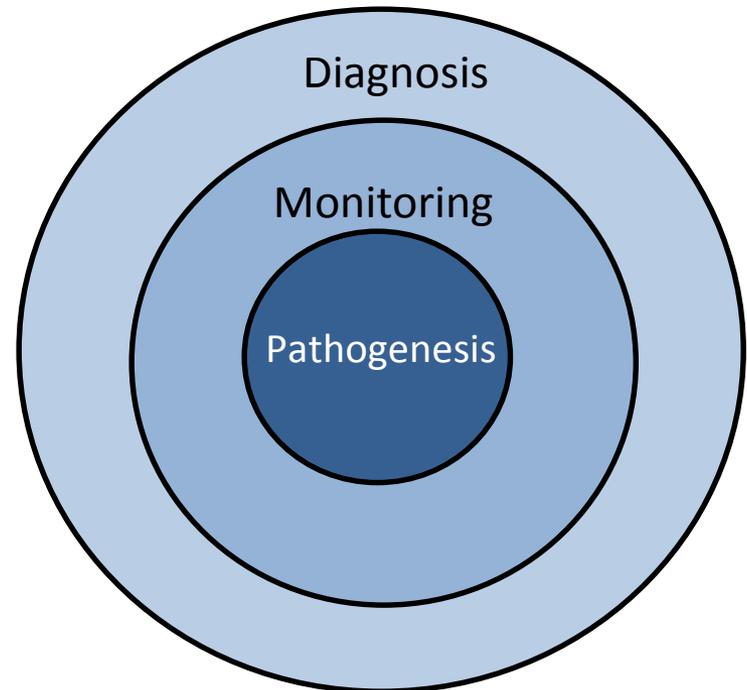
- Major investment by Oncologist to develop this technology for cancers
- Now used extensively for monitoring cancer patients during clinical trials and also for patient care
- ***Can these technologies be used in Infectious Diseases?***

# How can imaging help with TB trials and patients?

- Tomographic imaging can evaluate disease processes deep within the body, noninvasively and relatively rapidly.
- Longitudinal assessments can be conducted in the same individual, which is a fundamental advantage over the traditional invasive tools.
- Provides holistic, 3D views of the whole organ or body representative of the overall disease, and also less prone to sampling errors.

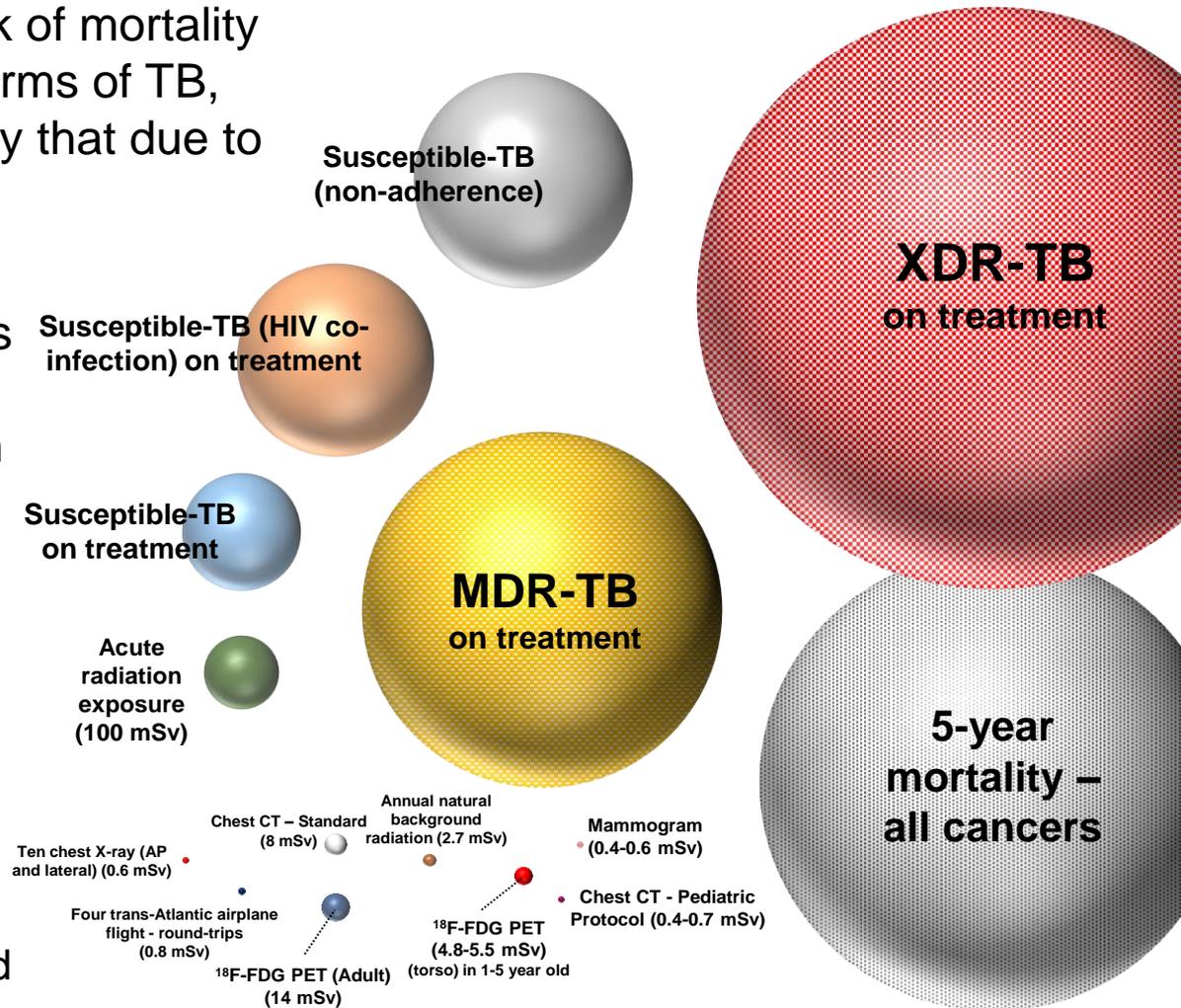
## Purpose

- Diagnosis
- Monitoring and prognostication;  
end-points for treatment trials  
enable adaptive designs
- Understanding pathogenesis  
host-directed therapies  
multi-compartment PK



# Risk of Imaging: Let's be real . . .

- Even with treatment, the risk of mortality is high for XDR and MDR forms of TB, and similar to 5-year mortality that due to cancers.
- Even drug-susceptible TB with adequate treatment has substantially higher risk of mortality (~1000 times) than radiation induced cancers with optimized imaging.
- Radiation due to:
  - Optimized PET (Pediatric torso), similar to annual background radiation
  - Chest CT – Pediatric protocol, similar to a mammogram or four round trip trans-Atlantic flights



# Following Cavity formation and Treatment Failure using High-Resolution CT Imaging

Development of a cavity during treatment failure, due to inadequate TB treatment

TB treatment

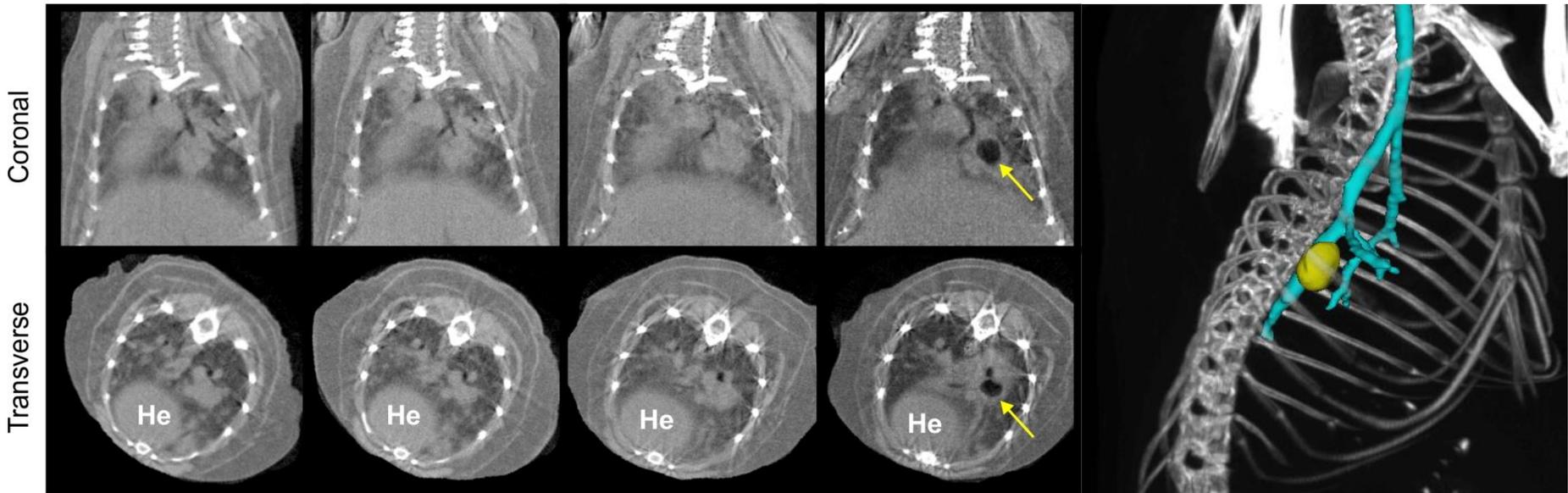
Failure

Week 9

Week 14

Week 20

Week 26



He = Heart

# TB granulomas (tumors) in mouse lungs

ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, Nov. 2009, p. 4879–4884  
0066-4804/09/\$12.00 doi:10.1128/AAC.00789-09  
Copyright © 2009, American Society for Microbiology. All Rights Reserved.

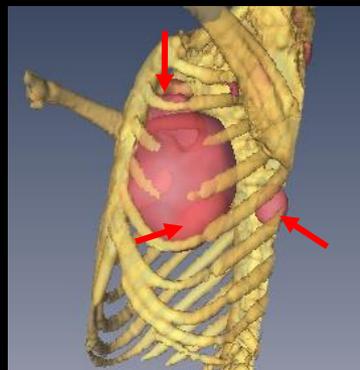
Vol. 53, No. 11

## Noninvasive Pulmonary [ $^{18}\text{F}$ ]-2-Fluoro-Deoxy-D-Glucose Positron Emission Tomography Correlates with Bactericidal Activity of Tuberculosis Drug Treatment $\nabla$ †

Stephanie L. Davis,<sup>1,2</sup> Eric L. Nuermberger,<sup>1,3</sup> Peter K. Um,<sup>1,2</sup> Camille Vidal,<sup>5,7</sup> Bruno Jedynak,<sup>6,7</sup> Martin G. Pomper,<sup>4</sup> William R. Bishai,<sup>1,3</sup> and Sanjay K. Jain<sup>1,2\*</sup>

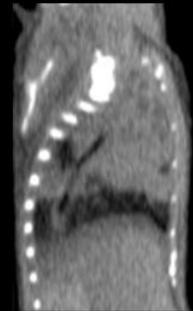
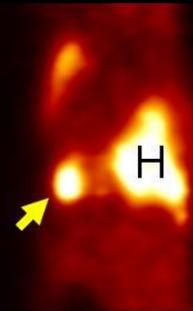
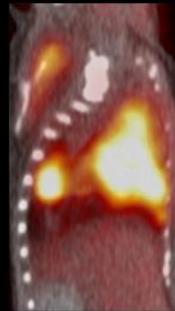
*Center for Tuberculosis Research,<sup>1</sup> Department of Pediatrics,<sup>2</sup> Department of Medicine,<sup>3</sup> and Department of Radiology,<sup>4</sup> Johns Hopkins University School of Medicine, Baltimore, Maryland 21287, and Department of Biomedical Engineering,<sup>5</sup> Department of Applied Mathematics and Statistics,<sup>6</sup> and Center for Imaging Science,<sup>7</sup> Johns Hopkins University, Baltimore, Maryland 21218*

Received 12 June 2009/Returned for modification 15 July 2009/Accepted 26 August 2009



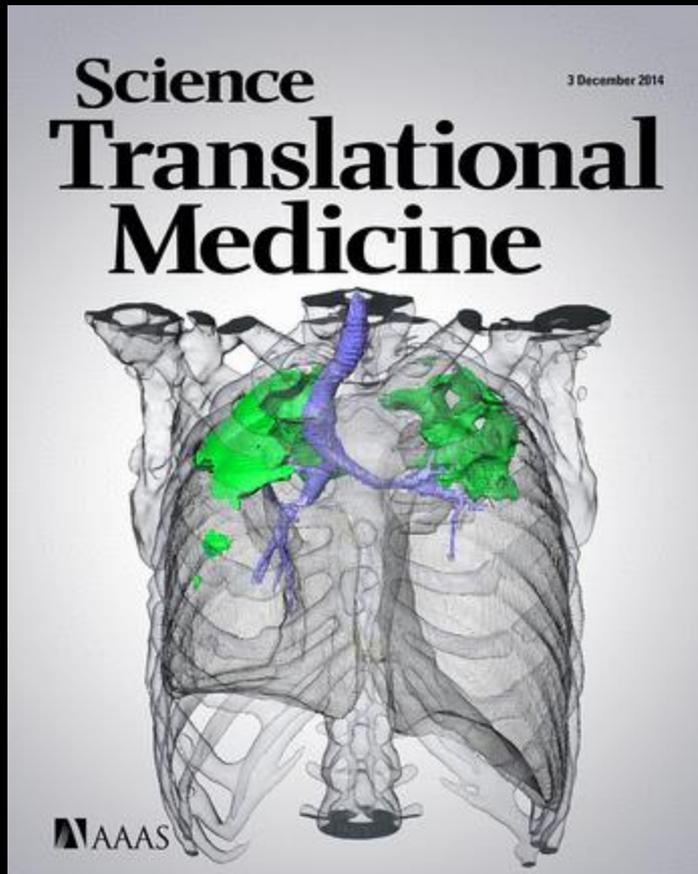
Sagittal

H = Heart



**Human data:** Sathekge *et al.* *J Nucl Med.* 2011;  
Chen *et al.* *Sci Transl Med.* 2014

# $^{18}\text{F}$ -FDG PET/CT correlates with treatment outcome in patients with MDR-TB

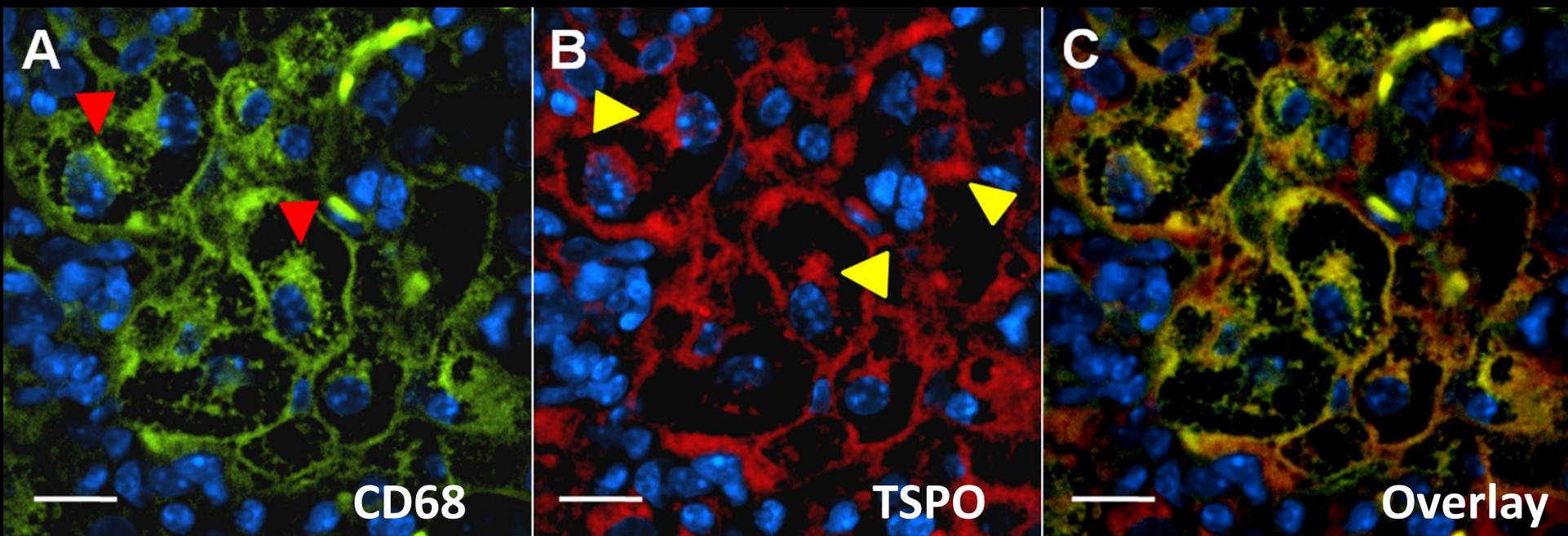


- Prospective imaged 35 adults (median age 37 years) with MDR-TB, on second-line TB treatment, using  $^{18}\text{F}$ -FDG PET and CT at 2 and 6 months after starting treatment .
- Imaging assessed by radiologists or automated analyses.
- $^{18}\text{F}$ -FDG PET at 2 months and automated CT at 6 months were more sensitive than sputum smear or solid culture conversion at 2 months, these differences were not statistically significant, possibly because of the small sample size in our study.
- Automated methods were more reliable than radiologists.

# Imaging TB-associated inflammation with iodo-DPA-713

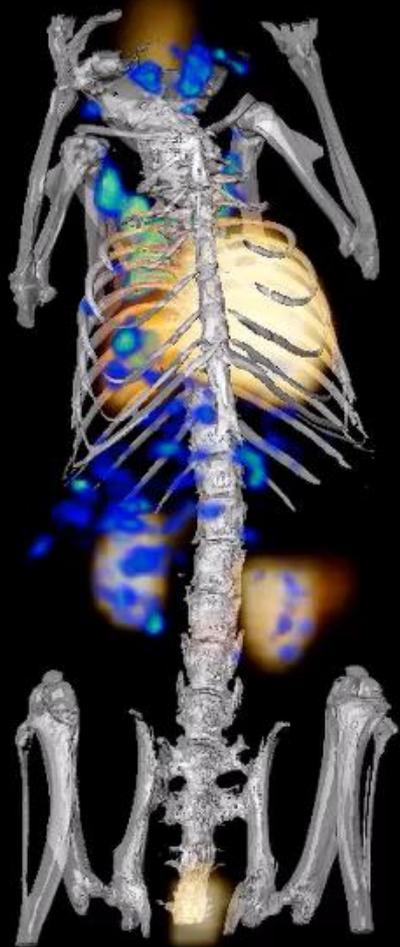
## DPA-713

- Iodo-DPA-713 is a ligand for translocator protein (TSPO)
- Up-regulated in inflamed microglia and macrophages
- TB lesions full of activated macrophages



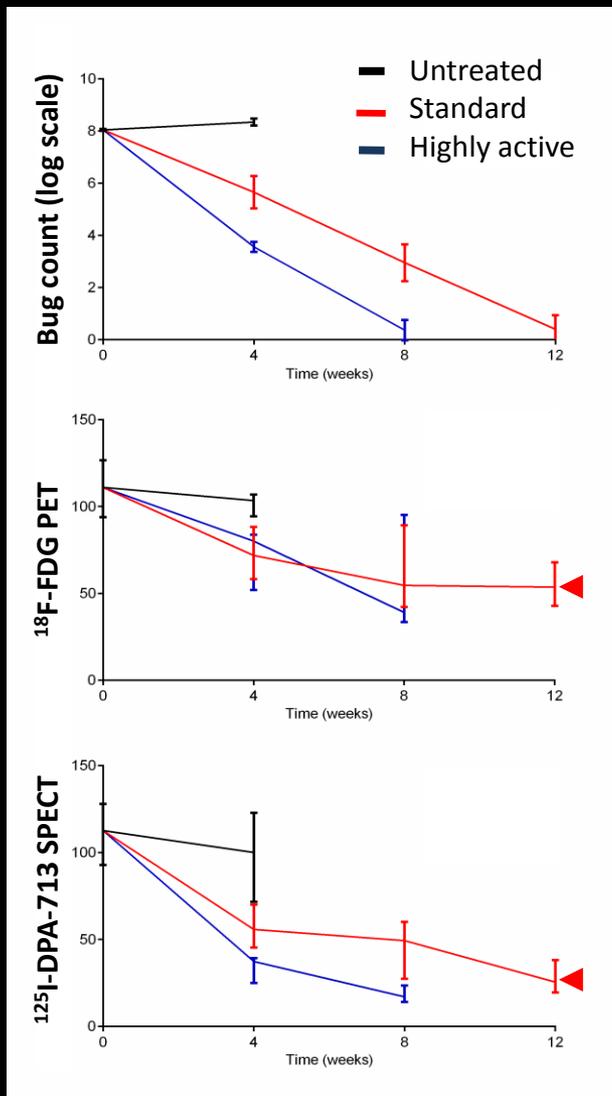
TSPO expression in macrophages within TB lesions

# Imaging TB-inflammation to monitor treatments: $^{125}\text{I}$ -DPA-713-SPECT versus $^{18}\text{F}$ -FDG-PET



$^{125}\text{I}$ -DPA-713-SPECT (blue-green)  $4.06 \pm 0.52$   
 $^{18}\text{F}$ -FDG-PET (orange)  $2.00 \pm 0.28$

skeletal outline (grey)



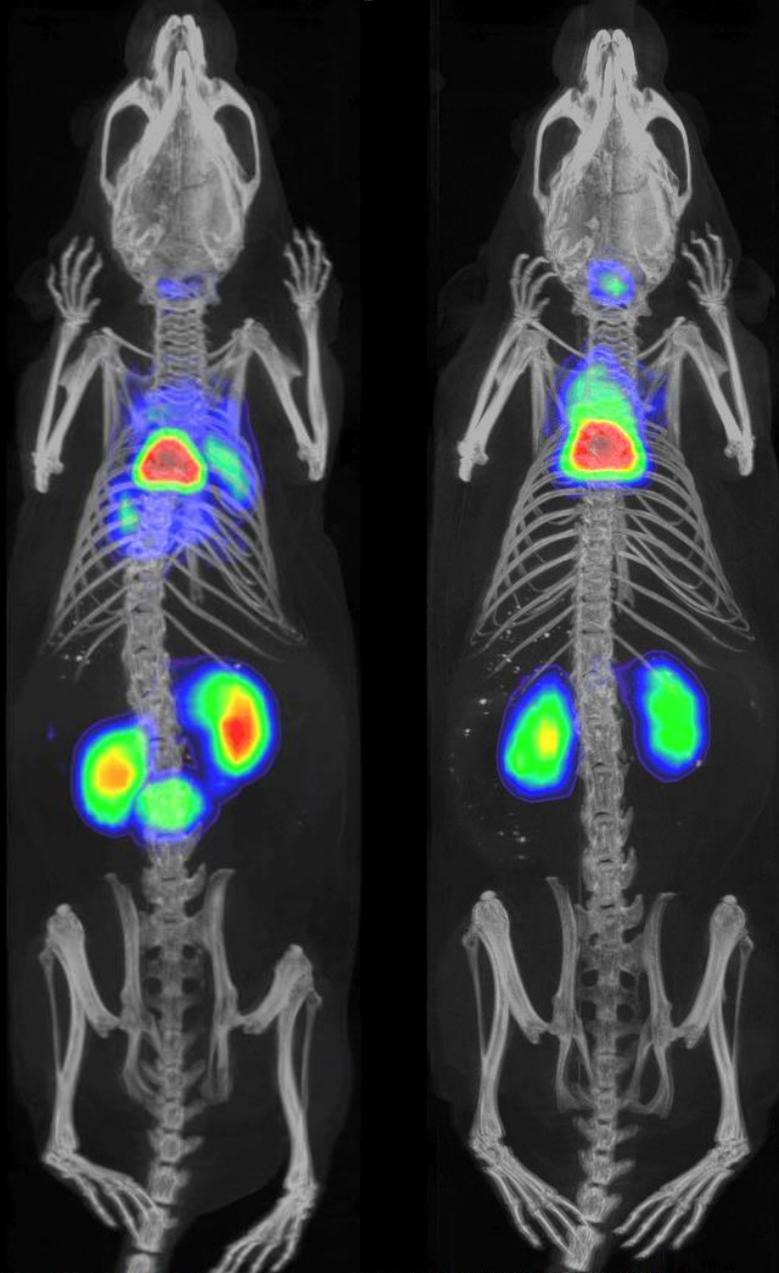
Pulmonary  $^{125}\text{I}$ -DPA-713 SPECT, *but not*  $^{18}\text{F}$ -FDG PET, correctly identified the bactericidal activities of the TB treatments as early as 4 weeks after starting treatment ( $P < 0.03$ )

Iodo-DPA-713 bound activated ( $\text{CD68}^+$ ) antigen presenting cells and imaging correlated with tissue  $\text{TNF-}\alpha$  (Spearman's  $\rho = 0.94$ ;  $P < 0.01$ )

Significant correlation was found between an increase in  $^{125}\text{I}$ -DPA-713 SPECT activity (*but not with*  $^{18}\text{F}$ -FDG PET) with bacterial burden at relapse (Spearman's  $\rho = 0.79$ ;  $P < 0.01$ )

# Imaging TB-associated inflammation with $^{124}\text{I}$ -DPA-713-PET

- $^{124}\text{I}$ -DPA-713 synthesized under a research contract
- Mouse studies demonstrate ( $^{124}\text{I}$ -DPA-713 PET in blue-green-red; and skeletal outline in grey) excellent signal-to-noise ratio: 4.0-fold higher in TB lesions versus uninfected controls (P = 0.03)
- Toxicology studies completed and first-in-human studies have started under an FDA Exploratory Investigational New Drug application (#121615)
- ***Half life of I-124 is 4.2 days and so this can be shipped worldwide.***



Jain\_AO\*H24vsI125\_Infected\_M5\_I124\_071114

0\*Jain\_AO\*H24vsI125\_Uninfected\_M7\_I124\_071114

0\*

Infected

Uninfected

# Proposed $^{124}\text{I}$ -DPA-713 PET human studies to specifically localize sites of TB infections

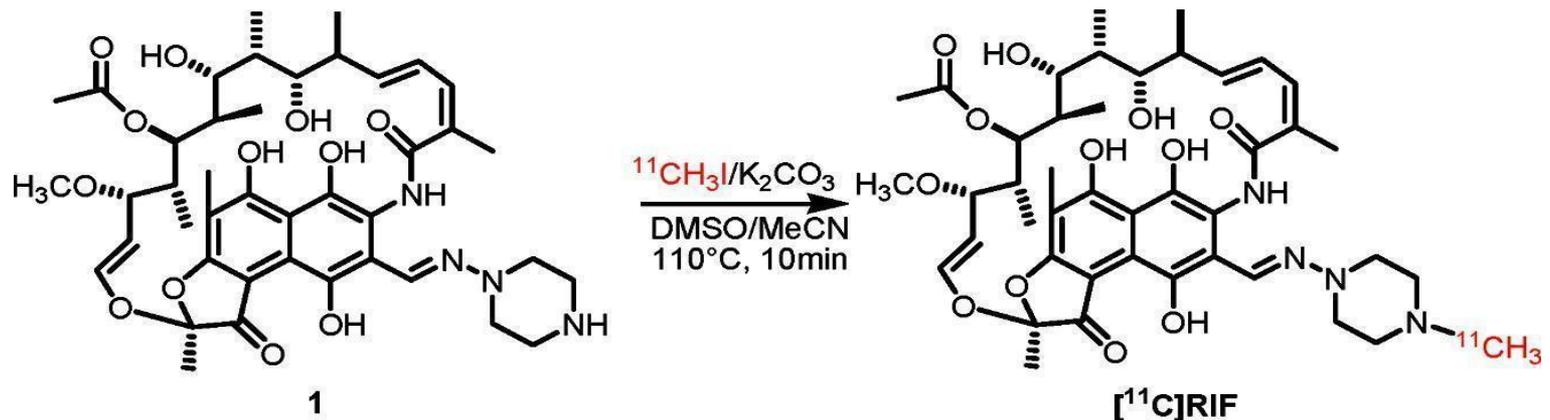
- Age: 18-65 years old
- Culture or molecular (GeneXpert, etc.) confirmation of TB
- On TB treatment for  $\leq 4$  weeks by the time of study
- Can understand English

If so, fill the screening script and refer to the study staff (JHU). The study, funded by the U.S. National Institutes of Health, requires three visits:

- Screening visit: history, written consent, screening labs
- Imaging visit day 1 (~half a day); injection of tracer
- Imaging (day 2 or 3) may last up to 3 hours
- During the imaging visit, we will use a new investigational PET/CT imaging technique
- Paid a total of \$200 for participation in this study

# Radiosynthesis and Bioimaging of Rifampin

- First-line drug essential for shortening therapy against *M. tuberculosis*
- Dosing based on serum / plasma concentrations (confirmed by post mortem resection)
- Drug concentration within necrotic pulmonary lesions (post-mortem) lower than blood concentrations\*
- Labeled rifampin with  $^{11}\text{C}$  using methods described by Liu *et al*

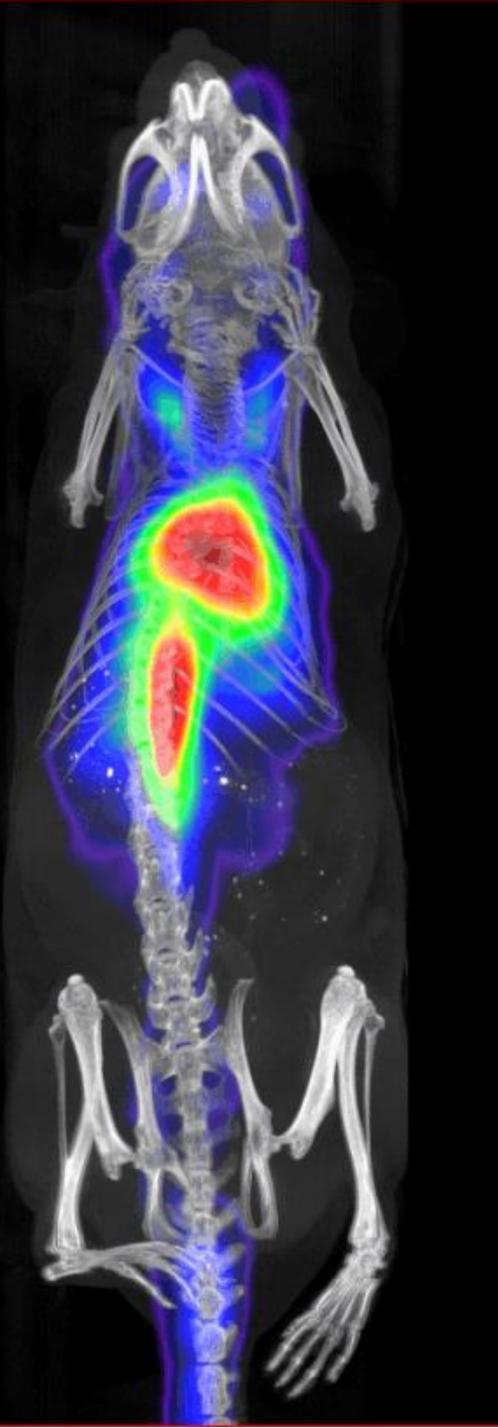


\*Kjellsson *et al. Antimicrob Agents Chemother.* 2012

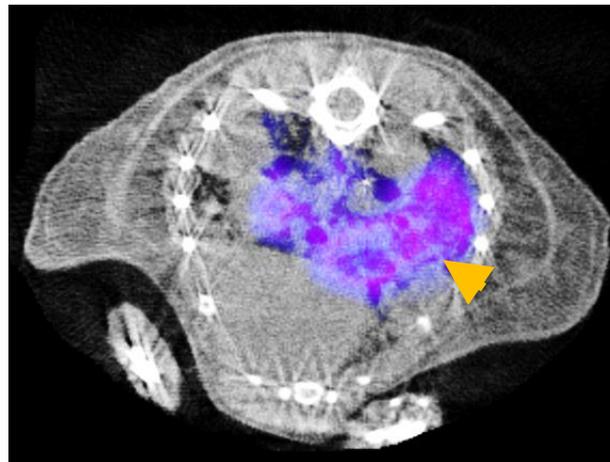
Liu *et al. J Med Chem.* 2010

# Radiosynthesis and Bioimaging of Rifampin

- Dynamic  $^{11}\text{C}$ -Rifampin PET/CT of a *M. tuberculosis* infected mouse
- Granulomatous tissue depicted by yellow circle
- Purple represents concentration of rifampin (highlighted by orange arrow)

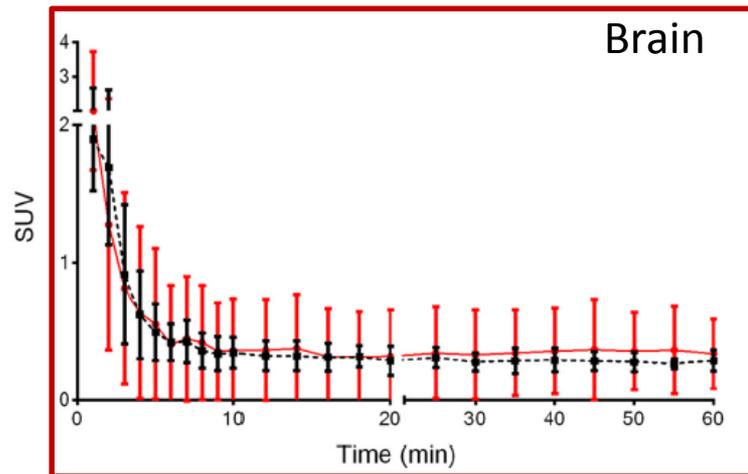
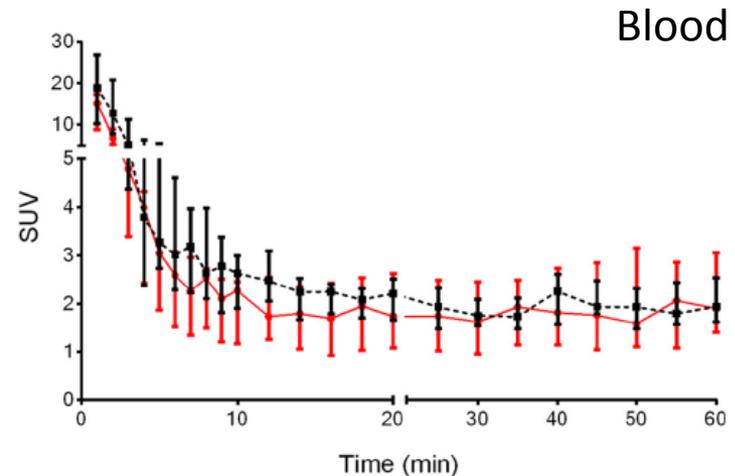
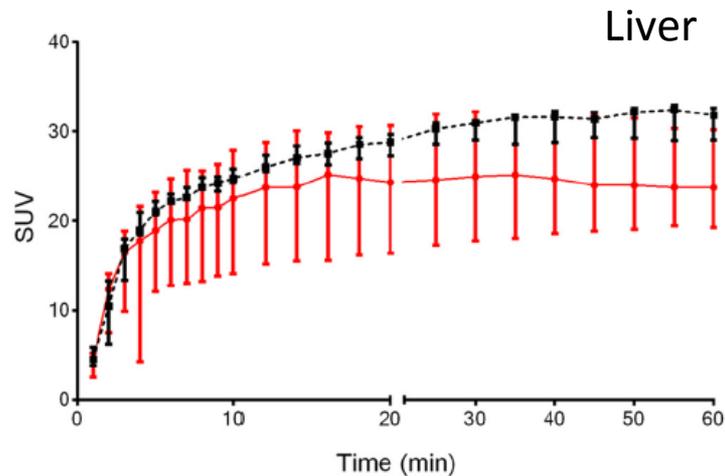


$^{11}\text{C}$ -Rifampin PET/CT

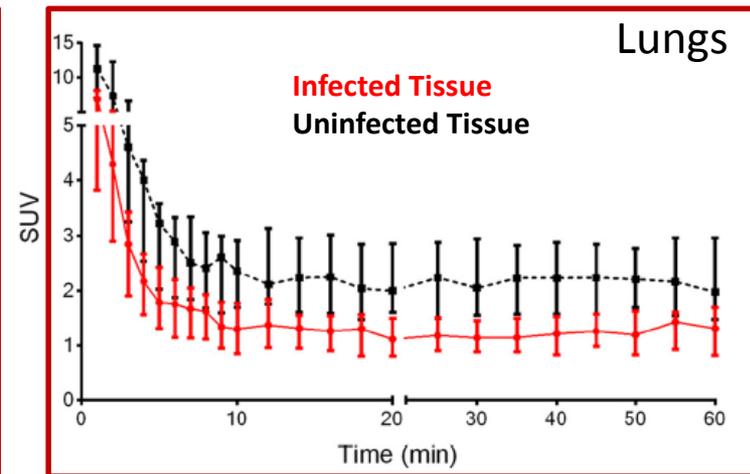


# Radiosynthesis and Bioimaging of Rifampin

## Lower concentrations in infected lung tissues



**$^{11}\text{C}$ -rifampin concentrations in the brain ~10-20% of blood levels**



**$^{11}\text{C}$ -rifampin concentrations were significantly lower in infected (~50%) versus uninfected lung tissues**

# Radiosynthesis and Bioimaging of Rifampin

## *Lower concentrations in infected lung tissues*

TABLE 1 Characteristics of study animals and results of noncompartmental pharmacokinetic analyses<sup>a</sup>

Parameter	Mean ± SD for animal type:		P value
	<i>Mycobacterium tuberculosis</i> infected	Uninfected	
Weight (g)	30.54 ± 1.90	34.18 ± 2.50	0.07
Injected dose (ng)	0.07 ± 0.02	0.07 ± 0.02	0.80
Injected dose (MBq)	8.61 ± 2.09	8.26 ± 2.51	0.80
Blood			
$C_{max}$ (ng/ml)	0.0622 ± 0.0289	0.0591 ± 0.0447	0.90
AUC <sub>0-60</sub> (ng · h/ml)	0.0056 ± 0.0015	0.0061 ± 0.0021	0.68
Brain			
$C_{max}$ (ng/ml)	0.0086 ± 0.0085	0.0050 ± 0.0022	0.39
AUC <sub>0-60</sub> (ng · h/ml)	0.0012 ± 0.0013	0.0008 ± 0.0025	0.53
Liver			
$C_{max}$ (ng/ml)	0.0608 ± 0.0167	0.0656 ± 0.0156	0.65
AUC <sub>0-60</sub> (ng · h/ml)	0.0583 ± 0.01406	0.0538 ± 0.0168	0.65
Lung			
$C_{max}$ (ng/ml)	0.0221 ± 0.0095	0.0326 ± 0.0205	0.33
AUC <sub>0-60</sub> (ng · h/ml)	0.0034 ± 0.0008	0.0054 ± 0.0015	0.03

<sup>a</sup> Rifampin concentrations (nanograms per milliliter) derived from PET imaging of *Mycobacterium tuberculosis*-infected and uninfected mice were analyzed using a noncompartmental intravenous bolus model (WinNonlin Standard). Peak concentrations ( $C_{max}$ ) and areas under the concentration-time curve for the first 60 min (AUC<sub>0-60</sub>) for different compartments are shown as means ± standard deviations. P values were calculated using a two-tailed Student *t* test. Five animals were used for each group.

**\*Human studies:** Dynamic <sup>11</sup>C-rifampin PET imaging studies in TB patients have begun to evaluate the penetration of rifampin in target, infected tissues – necrotic, cavitory and CNS lesions. **cGMP syntheses complete and RDRC / IRB approvals obtained.**

# Proposed $^{11}\text{C}$ -rifampin PET human studies in patients with Rifampin-susceptible TB

- Age:  $\geq 18$  years old
- Culture or molecular (GeneXpert, etc.) confirmation of TB susceptible to rifampin
- On TB treatment for  $\leq 6$  weeks by the time of study
- Can understand English

If so, fill the screening script and refer to the study staff (JHU). The study, funded by the U.S. National Institutes of Health, requires two visits:

- Screening visit: history, written consent, screening labs
- Imaging visit day 1 (~half a day)
- During the imaging visit, we will use a new investigational PET/CT imaging technique
- Paid a total of \$150 for participation in this study

# Current Imaging Agents: Lack of Infection-specific tracers

## Pros

Readily  
available

CT / MRI / US

## Cons

Non-specific for  
bacteria or  
infection

Excellent  
sensitivity

$^{18}\text{F}$ -FDG PET/ WBC scan

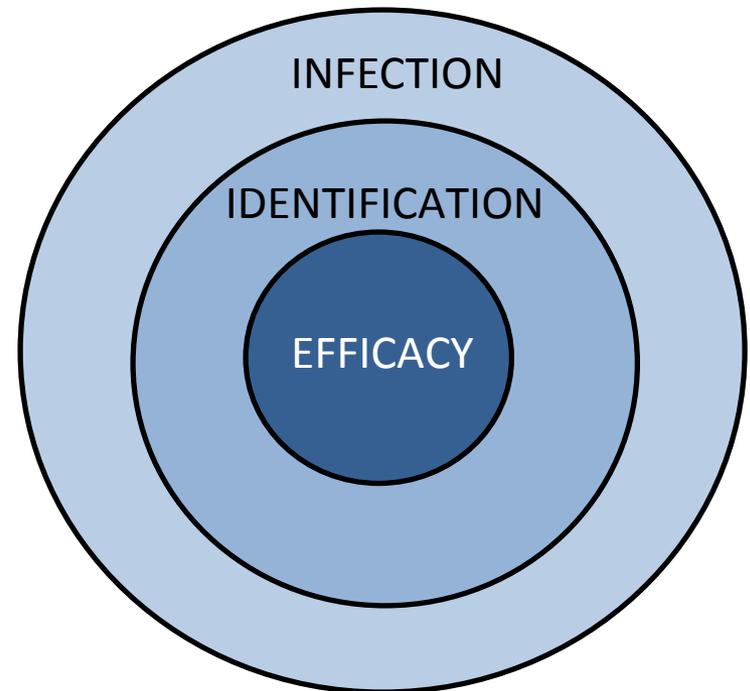
Non-specific;  
host-dependent

# Developing Bacteria-specific Imaging Tracers

**AIM:** To develop a pipeline of class specific bacterial imaging probes that would provide a platform to identify, localize and monitor a wide range of pathogenic bacteria

**HYPOTHESIS:** Small molecules metabolized by prokaryotic-specific pathways could be utilized as bacteria-specific imaging tracers to:

- a) discriminate infection from non-infectious processes
- b) categorize the causative bacterial species and
- c) provide information on antibiotic efficacy



# Candidate Bacteria-specific Imaging Tracers

Name	<i>In vitro</i> % uptake (mean ± standard deviation)				
	<i>S. aureus</i> (Gram-positive)	<i>E. coli</i> (Gram-negative)	<i>P. aeruginosa</i>	Mycobacteria* (Mtb)	Macrophages (J774)
L-Arabinose [1- <sup>14</sup> C]	0.41 ± 0.03	41.61 ± 9.91	0.21 ± 0.02	0.28 ± 0.01 (Mtb)	0.18 ± 0.01
Cellobiose [ <sup>3</sup> H]	1.81 ± 0.10	0.80 ± 0.05	--	0.13 ± 0.02 (Ms)	--
D-Lyxose [1- <sup>14</sup> C]	0.03 ± 0.01	1.86 ± 0.14	0.12 ± 0.04	0.35 ± 0.08 (Mtb)	0.04 ± 0.01
D-Mannitol [1- <sup>14</sup> C]	68.40 ± 7.39	81.80 ± 1.96	0.69 ± 0.05	0.29 ± 0.13 (Mtb)	0.12 ± 0.01
Methyl-α-D-glucopyranoside [methyl- <sup>14</sup> C]	11.01 ± 0.71	26.78 ± 0.59	--	0.11 ± 0.01 (Ms)	--
para-Aminobenzoic acid [3,5- <sup>3</sup> H]	16.82 ± 1.03	18.99 ± 5.80	4.02 ± 1.11	32.93 ± 4.73 (Mtb)	0.11 ± 0.01
L-Rhamnose [ <sup>3</sup> H]	4.96 ± 0.13	4.73 ± 0.07	0.24 ± 0.04	3.82 ± 0.84 (Mtb)	0.60 ± 0.01
Shikimic acid [3- <sup>3</sup> H]	7.54 ± 0.01	1.52 ± 0.02	1.31 ± 0.02	0.17 ± 0.01 (Ms)	--
D-Sorbitol [ <sup>14</sup> C] ( <sup>18</sup> F-FDS) <sup>†</sup>	0.47 ± 0.09	72.20 ± 9.09	0.52 ± 0.46	--	0.21 ± 0.01
D-Xylose [1- <sup>14</sup> C]	0.31 ± 0.01	73.94 ± 2.06	0.53 ± 0.08	0.18 ± 0.02 (Mtb)	0.19 ± 0.01

Detect the presence of bacterial infection

Identify the "type" of bacteria

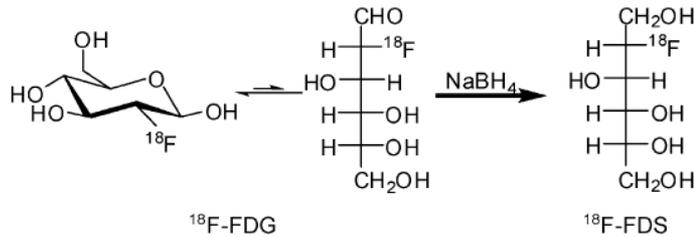
**FDS or Fluorodeoxysorbitol is a fluoro analog of Sorbitol – FDA approved sugar-free sweetener**



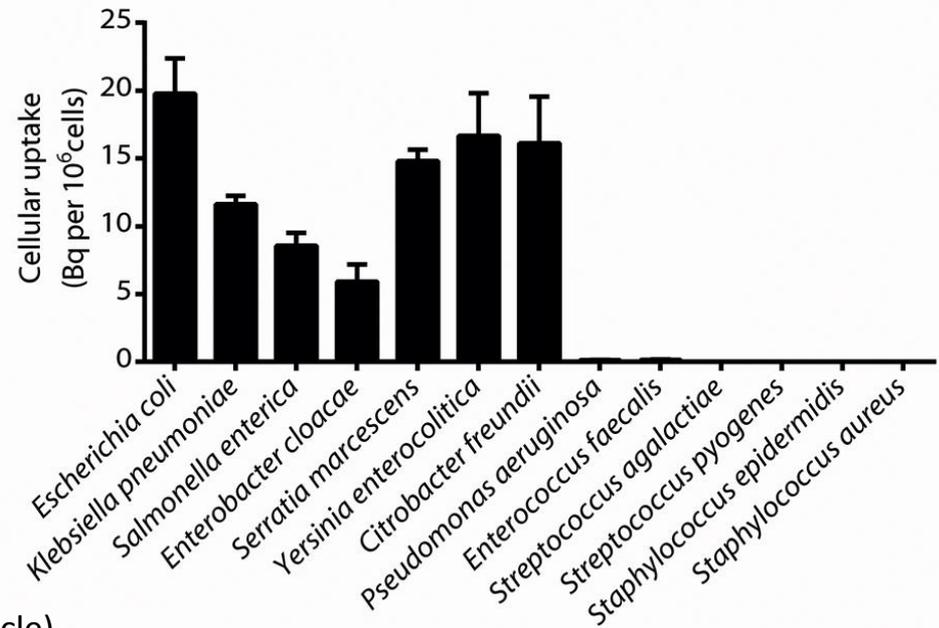
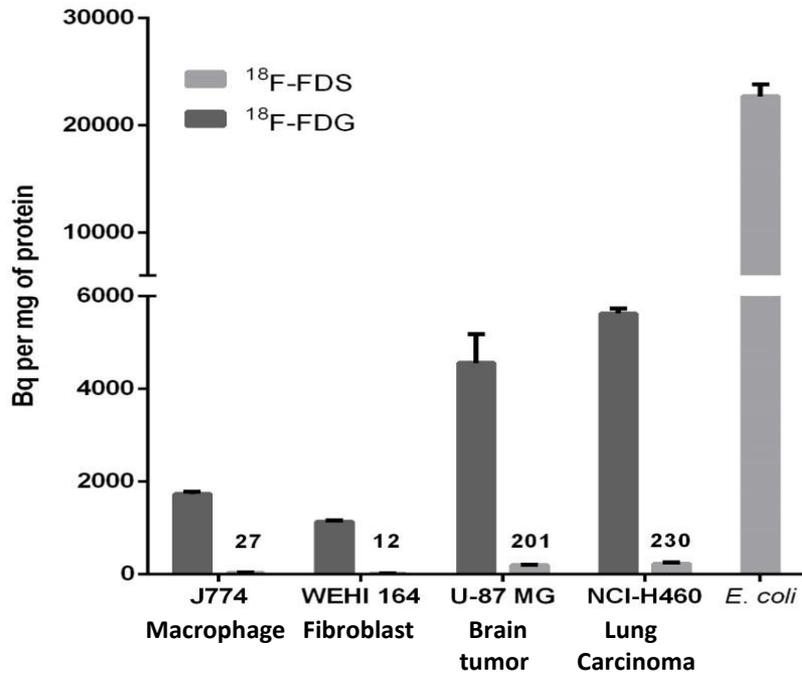
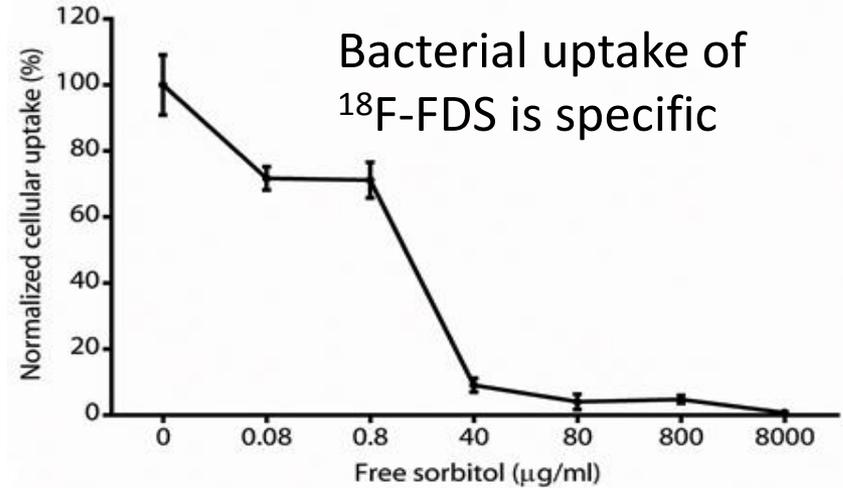
\**Mycobacterium smegmatis* = Ms or *Mycobacterium tuberculosis* = Mtb  
<sup>†</sup> 2-[<sup>18</sup>F]-fluorodeoxysorbitol (<sup>18</sup>F-FDS) used for uptake assays

**No uptake by heat killed bacteria for any of the tracers demonstrating specificity**

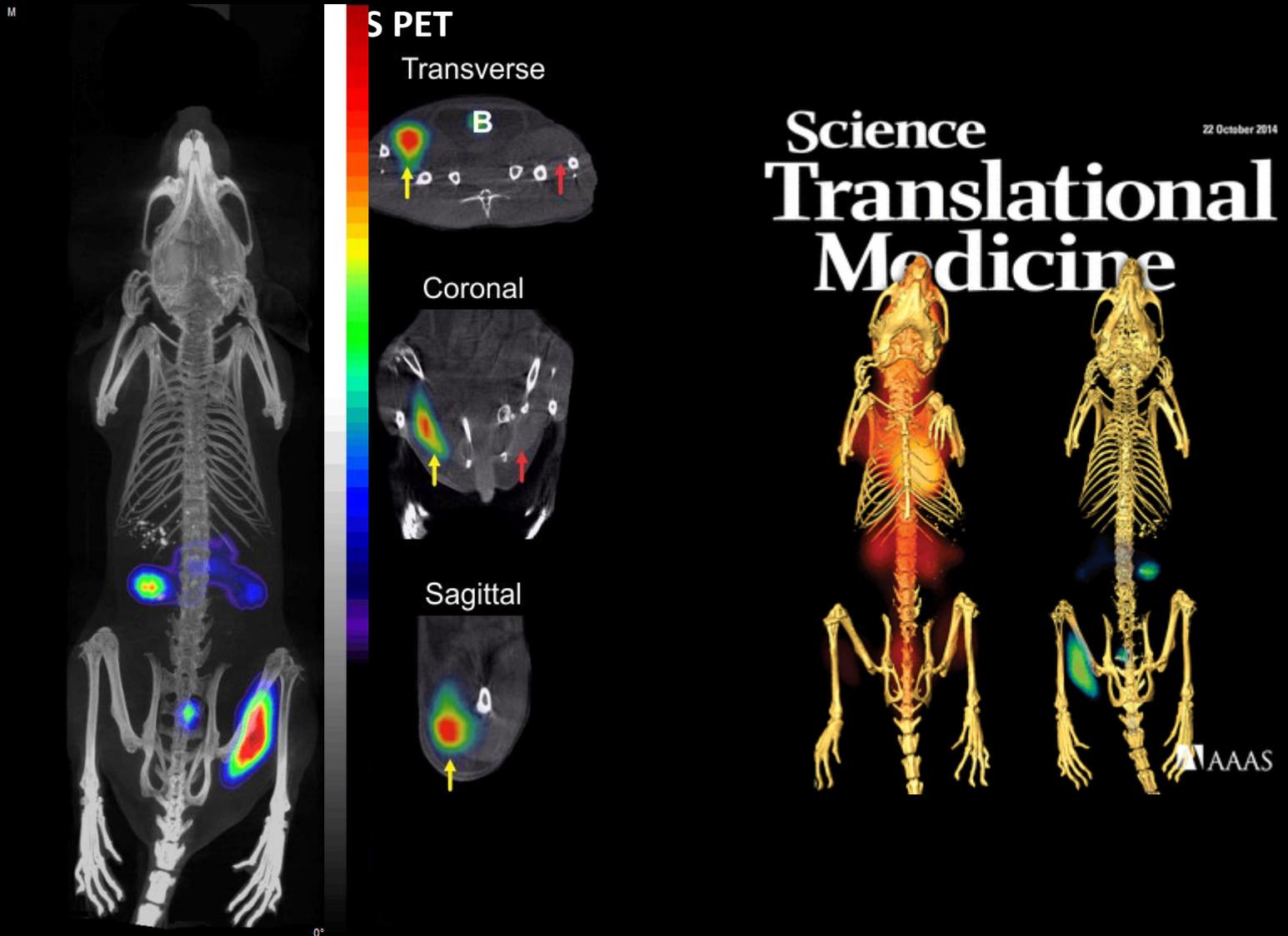
# $^{18}\text{F}$ -FDS can be synthesized from $^{18}\text{F}$ -FDG (in 30 min) and is specifically accumulated by *Enterobacteriaceae*



Li et al. *Mol Imaging Biol* 2008

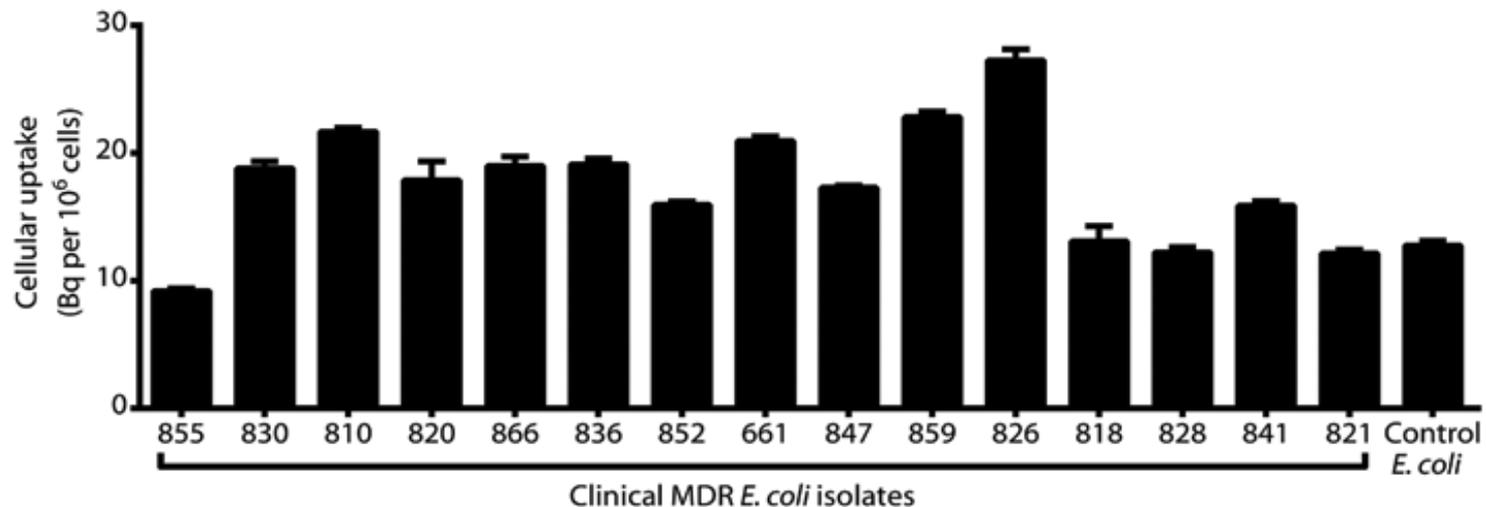


# $^{18}\text{F}$ -FDS PET can differentiate infection sites from sterile inflammation

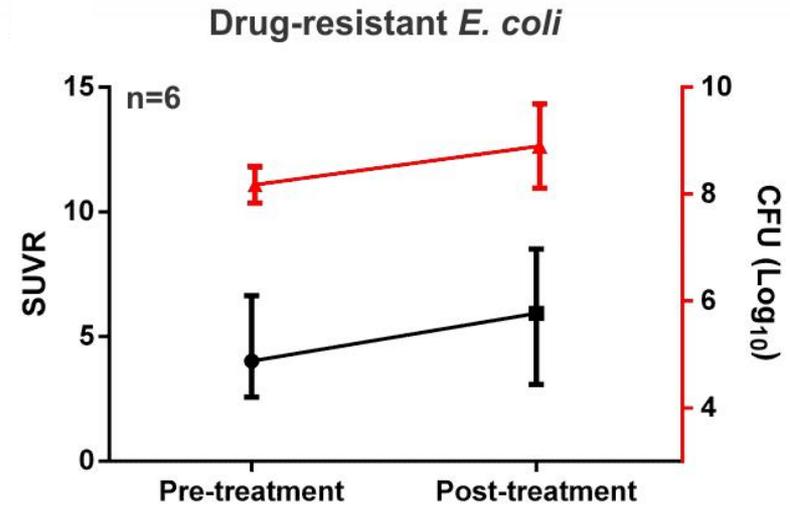
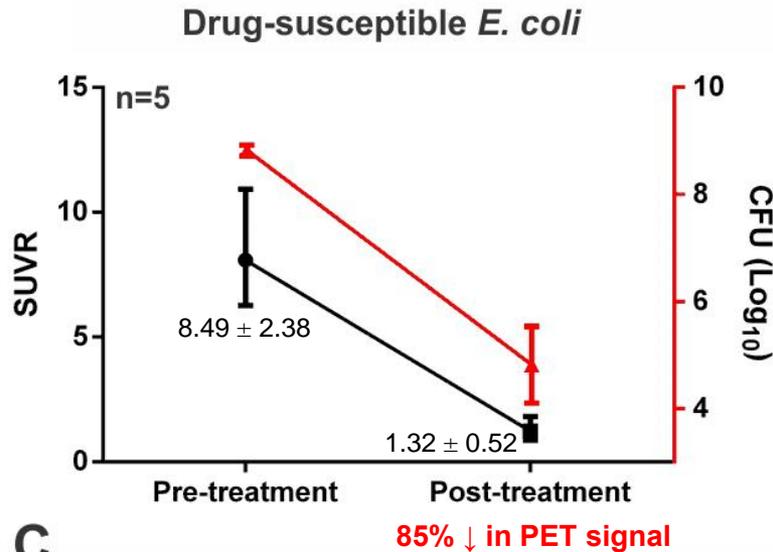


# $^{18}\text{F}$ -FDS Uptake by 15 Random Clinical Multidrug-Resistant *Enterobacteriaceae* strains

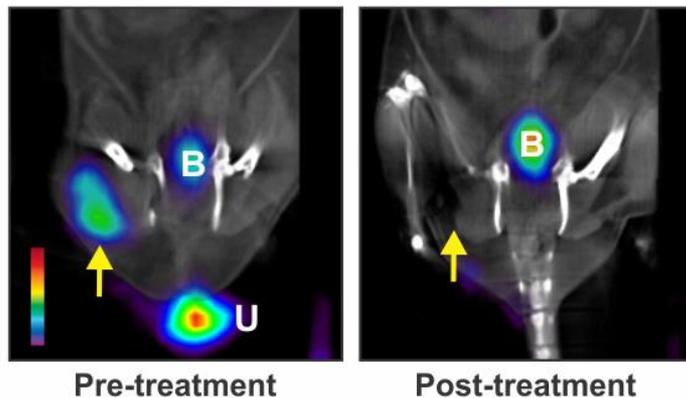
- Notorious source of life-threatening nosocomial infections due to multi-drug resistant organisms including:
  - Extended-spectrum beta-lactamase (ESBL)-producing *Enterobacteriaceae*
  - Carbapenem-resistant *Enterobacteriaceae* (CRE)
  - *Enterobacteriaceae* resistant to colistin (“last line of defense”)
- All 15 random ESBL-producing clinical strains (clinical MDR *E. coli*), demonstrated substantial  $^{18}\text{F}$ -FDS uptake



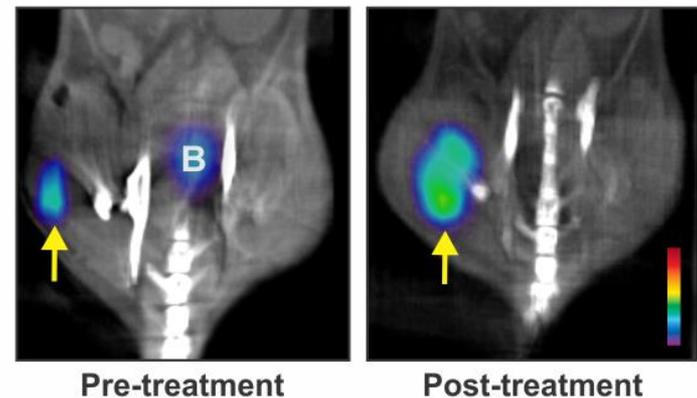
# Monitoring antimicrobial efficacy in multidrug-resistant infections *in situ*



C



D



An increase in bacterial load corresponds with disease progression to sepsis and death in seriously ill patients

## XDR-TB report

### JHU

Nicole Salazar-Austin  
Alvaro Ordonez  
Alice Jenh Hsu  
Jane Benson  
Mahadevappa Mahesh  
Aaron Milstone  
Nicole Parrish  
Eric Nuermberger  
Richard Chaisson  
Nursing and Pharmacy

### Others

Jeffrey Starke  
Max Salfinger

### Public Health

**Nancy Baruch**  
Elizabeth Menachery  
Jafar Razeq  
Dorothy Freeman

Maryland Department of Health and Mental Hygiene  
Howard County Health Department  
Centers for Disease Control and Prevention  
Food and Drug Administration

### Jain Lab

Alvaro Ordonez  
Ed Weinstein  
Supriya Pokkali  
Liz Tucker  
Alvin Kalinda  
Vikram Saini  
Mariah Klunk  
Peter DeMarco  
Lauren Bambarger  
Yongseok Chang  
Allison Murawski

### Stony Brook

Peter Tonge  
Hui Wang  
Zhang Zhuo

# Acknowledgments



### JHU Radiology

Catherine Foss  
Ronnie Mease  
Martin Pomper  
Bob Dannals  
Steve Rowe  
Akimosa Jeffrey-  
Kwanisai  
Ghedem Solomon  
Mahadevappa  
Mahesh

### JHU

Carlton Lee  
Maunank Shah  
Kelly Dooley

### Maryland TB Counties

Kelly Russo  
Elizabeth Menachery  
Lucia Donatelli  
Kimberly Townsend

## Funding

*NIH Director's Transformative Research Award R01-EB020539*

*NIH Director's New Innovator Award DP2-OD006492*

*NHLBI R01-HL116316, R01-HL131829 and NIAID DAIDS supplement*

