

TB Cross-Contamination from the laboratory perspective

Jafar H Razeq, Ph.D., HCLD (ABB)
Chief, Division of Microbiology
Laboratories Administration
State of Maryland DHMH
Jafar.razeq@Maryland.gov

The J. Mehsen Joseph Public Health Laboratory



New Home

State of Maryland

Department of Health and Mental Hygiene

Laboratories Administration

1770 Ashland Ave
Baltimore, Md. 21205



TB Cross-Contamination

“False-Positive Cultures”

Consequences of false positive cultures:

- First and foremost: exposes the patient to unnecessary, and potentially toxic treatment, hospitalization, isolation, and anxiety
- Unnecessary follow-up and contact investigation
- Unnecessary laboratory tests and work
- Potential legal aspects

- Rates of false positive cultures has been reported to be ranging from 2.2%-10.5%
- Most common causes of false positive cultures are clerical errors, contamination of clinical equipment, and laboratory cross-contamination
- Laboratory cross-contamination is the mostly described and documented

Causes of False-Positive Cultures

I. CLERICAL ERRORS: MISLABELING

The clerical errors have been associated with mislabeling of specimens leading to a false-positive culture result.

There is limited data on this issue.

Causes of False-Positive Cultures

II. CONTAMINATION OF CLINICAL EQUIPMENT

Contamination of clinical equipment (e.g., bronchoscope) can cause false-positive culture.

A contaminated device can cause both false-positive cultures and tuberculosis transmission.

Causes of False-Positive Cultures

III. LABORATORY CROSS-CONTAMINATION

Our topic for today.

Definition of Laboratory Cross Contamination

The transfer of MTB complex bacilli from one specimen to another specimen that does not contain viable bacilli, causing a false-positive result.

Causes of Cross-Contamination ~ how it happens:

- No environmental reservoir for MTB, but it is hardy organism and can survive harsh conditions.
- Creation of aerosol or splashing during specimen processing
- Defects in the exhaust systems of the BSCs used for specimen processing. The creation of an aerosol when samples are processed that is not cleared promptly (through a functional BSC) may lead to the settlement of these particles in the subsequent specimen

Causes of Cross-Contamination ~ how it happens:

- Use of common flask to dispense reagents.
- Specimen carryover, spillage, or inadvertent contamination during specimen processing.
- Contamination of pipettes or lids.
- Batch processing of specimen is the efficient way of testing, but carries the potential for cross-contamination due to contamination common reagents like the buffer.



Causes of Cross-Contamination ~ how it happens (continued):

- Processing excessive number of specimens in one batch due to staff shortages may lead to breakdown in protocol adherence and introduce potential error for cross-contamination.

To reduce the possibility of Cross-Contamination:

- Use daily aliquots of processing reagents and buffers. Any leftover should be discarded.
- Never use common beakers or flasks when processing.
- Keep the specimen tubes tightly closed and clean the outside of the tube prior to vortexing or shaking.
- Pour decontamination reagents or buffers slowly on the side of the tube without causing any splashing. Do not touch the container of reagents to the lip of the tube at any time during addition.

To Reduce the Possibility of Cross-Contamination(continued):

- Only one tube should be uncapped at a time.
- After mixing or vortexing, wait for five (5) minutes before uncapping the tubes.
- Open the specimen tubes very gently to avoid aerosol generation.
- When adding reagents to the tube, open one tube at a time. Do not keep all the tubes open at the same times.

To Reduce the Possibility of Cross-Contamination (continued):

- Establishment of a threshold which, when exceeded, will prompt an investigation.
- Standardizing of laboratory procedures.
- Do not place tubes too close to each other in the rack.
- Change gloves often.
- Avoid manipulation of PT specimens.
- Disinfect Biological Safety Cabinet (BSC) work surfaces routinely.

Criteria (i.e., **red flags**) that might trigger false-positive culture investigation:

- *The problem is that some of these red flags are also seen in true TB patients*
- Patient's clinical course is inconsistent with TB and the clinician is considering an alternative diagnosis.
- Single positive *M. tuberculosis* culture with no AFB seen in any specimen (although this can also be seen in true TB patients).
- Time to growth detection is > 30 days (this can also be seen in true TB patients).
- Clustering of MTB isolates with unusual resistance pattern in a short period of time.

Criteria (i.e., **red flags**) that might trigger false~positive culture investigation (continued):

- If MTB is cultured from a sample processed together with a smear positive specimen.
- Usually, specimens of false positive cultures are negative for AFB smear.
- **Culture~positive specimen from a different patient processed or handled on the same day has an identical DNA fingerprint and no epidemiologic links exist between patients.**

Reporting Language

*“Molecular genotyping tests performed at a reference laboratory on this particular isolate show that cross-contamination with Mycobacterium tuberculosis (MTB) from an extraneous source cannot be ruled out. Please interpret the positive report of MTB from this particular specimen with caution. If clinically warranted, please resubmit another specimen for further testing. As always, laboratory results cannot replace or override clinical judgment in the **diagnosis and management** of this patient. If further consultation on this case is needed, please consult the State TB Program at 410-767-6698. For laboratory related questions, call 410-767-6130/6125”*

Bottom line:

Treat patients, not laboratory results”

References

Chest, 2013; 144(1):319-322

MMWR, 2011; 49(19):413-416

Chest, 2010; 137(5):1065-1070

EID, 2002; 8 (11); 1260-1263

CID, 2000; 31: 1390-1395

Diag Micro Inf Dis, 2012; 73: 343-349

Clin Micro Rev, 2006; 658-685

Internal Med J, 2002; 512-519

CID, 2004; 38(15): e53-e54

JCM, 2006; 44(8): 2967-2969

Clin Micro Inf Dis, 2006; 12(10): 1042-1045

Arch Pathol Lab Med, 2000; 124: 902-903