

SUSPECT MERS-CoV SPECIMEN SUBMISSION GUIDELINES MARYLAND DHMH LABORATORIES ADMINISTRATION

JUNE 25, 2013

I. ACCEPTABLE SPECIMENS

1. **Lower Respiratory Specimens:** Sputum, bronchoalveolar lavage, tracheal aspirate
2. **Upper Respiratory Specimens:** Nasopharyngeal and Oropharyngeal Swabs in viral transport media
3. **Stool**
4. **Serum**

II. SPECIMEN COLLECTION KIT INSTRUCTIONS

Local health departments can order the UTM specimen collection kit and leak-proof, screw cap by calling 410-767-6120 or by faxing a completed request form (<http://dhmh.maryland.gov/laboratories/docs/Request%20Form.pdf>) at 410-333-5019.

III. SPECIMEN COLLECTION

Source: Interim Guidelines for Collection, Processing and Transport of Clinical Specimens from Patients under Investigation for Middle East Respiratory Syndrome (MERS)
<http://www.cdc.gov/coronavirus/mers/downloads/Interim-Guidelines-MERS-Collection-Processing-Transport.pdf>.

1. Lower Respiratory Specimens

a. Sputum

Have the patient rinse the mouth with water and then expectorate deep cough sputum directly into a sterile, leak-proof, screw-cap sputum collection cup or sterile dry container.

b. Lower respiratory tract aspirates/washes (Bronchoalveolar lavage, tracheal aspirate)

Collect 2-3 mL into a sterile, leak-proof, screw-cap sputum collection cup or sterile dry container.

2. Upper Respiratory Samples: Nasopharyngeal (NPS) and Oropharyngeal swabs (OPS) in viral transport media (VTM).

Use only synthetic fiber swabs with plastic shafts. **Do not use calcium alginate** swabs or swabs with wooden shafts, as they may contain substances that inactivate some viruses and inhibit PCR testing. Place swabs immediately into sterile tubes containing 2-3 ml of viral transport media.

Nasopharyngeal swabs: Insert a swab into the nostril parallel to the palate. Leave the swab in place for a few seconds to absorb secretions. Swab both nostrils.

Oropharyngeal swabs: Swab the posterior pharynx, avoiding the tonsils and tongue.

3. Stool

Collect 2-5 grams of stool specimen (formed or liquid) in sterile, leak-proof, screw-cap.

4. Serum

Serum specimens should be collected during the acute stage of the disease, preferably during the first week after onset of illness, and again during convalescence, ≥ 3 weeks later.

Children and adults: Collect 1 tube (5-10 mL) of whole blood in a serum separator tube. Allow the blood to clot, centrifuge briefly, and separate sera into sterile tube container.

Infants: A minimum of 1 cc of whole blood is needed for testing of pediatric patients. If possible, collect 1 cc in an EDTA tube and in a serum separator tube. If only 1cc can be obtained, use a serum separator tube

III. LABORATORY TEST REQUEST SLIP (INFECTIOUS AGENTS: CULTURE/DETECTION) INSTRUCTIONS

See the MERS-CoV Test Requisition Sample Form for guidance.

IV. PACKAGING AND SHIPPING

If you suspect a MERS CoV infection, please contact your Local Health Department for a review of the case. If testing is indicated the Local Health Department will make arrangements with the DHMH Laboratory for courier pick-up of specimens and expedited MERS CoV testing.

For questions or concerns, please contact the Division of Molecular Biology Laboratory at (410)767-5819 during normal business hours from 8:00AM - 4:30PM Monday through Friday.

For urgent inquiries after normal business hours, please contact the DHMH Lab emergency contact number at (410-925-3121).

2-10675



P.O. Box 2355 • Baltimore, MD 21203-2355
410-767-6100 www.dhmh.state.md.us/labs
Robert A. Myers, Ph.D., Director

<input type="checkbox"/> EH <input type="checkbox"/> FP <input type="checkbox"/> DMT/PN <input type="checkbox"/> QND <input type="checkbox"/> STD <input type="checkbox"/> TB <input type="checkbox"/> CD <input type="checkbox"/> COR		Patient SS# (last 4 digits):	
Health Care Provider		Last Name <input type="checkbox"/> SR <input type="checkbox"/> JR <input type="checkbox"/> Other	
Address		First Name M.I. Maiden:	
City	County	Date of Birth (mm/dd/yyyy) / /	
State	Zip Code	Address	
Contact Name:		City	County
Phone#	Fax#	State	Zip Code
Test Request Authorized by:			
Sex: <input type="checkbox"/> Male <input type="checkbox"/> Female <input type="checkbox"/> Transgender M to F <input type="checkbox"/> Transgender F to M		Ethnicity: Hispanic or Latino Origin? <input type="checkbox"/> yes <input type="checkbox"/> no	
Race: <input type="checkbox"/> American Indian/Alaska Native <input type="checkbox"/> Asian <input type="checkbox"/> Black/African American <input type="checkbox"/> Native Hawaiian/other Pacific Islander <input type="checkbox"/> White			
Case #	DOC#	Outbreak #	Submitter Lab#
Collect Date:	Collect Time:	<input type="checkbox"/> am <input type="checkbox"/> pm	Onset Date:
Reason for Test: <input type="checkbox"/> Screening <input type="checkbox"/> Diagnosis <input type="checkbox"/> Contact <input type="checkbox"/> Test of Cure 2-3 Months Post Rx <input type="checkbox"/> Suspected Carrier <input type="checkbox"/> Isolate for ID <input type="checkbox"/> Release			
Therapy/Drug Treatment: <input type="checkbox"/> No <input type="checkbox"/> Yes		Therapy/Drug Type: Therapy/Drug Date:	

SPECIMEN CODE		SPECIMEN CODE		SPECIMEN CODE	
BACTERIOLOGY		SPECIAL BACTERIOLOGY		RESTRICTED TESTS	
Bacterial Culture - Routine		Legionella Culture		Pre-approved submitters only	
Additional specimen codes: _____		Leptospira		Chlamydia trachomatis/GC NAAT	
Bordetella pertussis		Mycoplasma		Chlamydia trachomatis only/NAAT	
Group A Strep		MYCOBACTERIOLOGY/AFB/TB		Norovirus ** (see comment on back)	
Group B Strep Screen		AFB/TB Culture and Smear		OTHER TESTS FOR INFECTIOUS AGENTS	
C. difficile Toxin		AFB/TB Referred Culture for ID		Test name: _____	
Diphtheria		M. tuberculosis Referred Culture for Genotyping		Prior arrangements have been made with the following DHMH Laboratories	
Foodborne Pathogens (B. cereus, C. perfringens, S. aureus)		Nucleic Acid Amplification Test for M. tuberculosis Complex (MTD)		Administrative employee: _____	
Gonorrhea Culture/Incubated? <input type="checkbox"/> Yes <input type="checkbox"/> No		PARASITOLOGY			
Hrs. incubated: _____ Add'l specimen codes: _____		Blood Parasites: _____			
MRSA (rule out)		Country visited outside US: _____			
VRE (rule out)		Ova & Parasites/Immigrant? <input type="checkbox"/> Yes <input type="checkbox"/> No			
ENTERIC INFECTIONS		Cryptosporidium		SPECIMEN CODE	
Campylobacter		Cyclospora/Isospora		PLACE CODE IN BOX NEXT TO TEST	
E. coli O157 typing		Microsporidium		B Blood	
Enteric Culture - Routine (Salmonella, Shigella, E. coli O157, Campylobacter)		Pinworm		BW Bronchial Washing	
Salmonella typing		VIRUS ISOLATION/CHLAMYDIA		CSF Cerebrospinal Fluid	
Shigella typing		Adenovirus*		CX Cervix/Endocervix	
V. parahaemolyticus		Arbovirus Panel (WNV, EEEV, SLEV)		E Eye	
Yersinia		Chlamydia trachomatis		F Feces	
REFERENCE MICROBIOLOGY		Cytomegalovirus (CMV)		N Nasopharyngeal nasal	
ABC'S (BIDS) # _____		Enterovirus (Inc. Echo & Coxsackie)		P Penis	
Organism: _____		Herpes Simplex Virus (Types 1 & 2)		R Rectum	
Bacteria Referred Culture for ID		Influenza (Types A & B)*		SP Sputum	
Specify: _____		Parainfluenza (Types 1, 2 & 3)*		T Throat	
		Respiratory Syncytial Virus (RSV)*		URE Urethra	
		Varicella (VZV)		UFV Urine (First Void)	
				UCC Urine (Clean Catch)	
				V Vagina	
				W Wound	
				O Other	

SPECIMEN CODE: _____
 PLACE CODE IN BOX NEXT TO TEST
 B Blood
 BW Bronchial Washing
 CSF Cerebrospinal Fluid
 CX Cervix/Endocervix
 E Eye
 F Feces
 N Nasopharyngeal/ nasal
 P Penis
 R Rectum
 SP Sputum
 T Throat
 URE Urethra
 UFV Urine (First Void)
 UCC Urine (Clean Catch)
 V Vagina
 W Wound
 O Other: _____

1. lower respiratory specimen:
sputum, bronchoalveolar lavage, tracheal aspirate
2. upper respiratory specimen:
nasopharyngeal & oropharyngeal swabs in VTM
3. stool
4. serum

For additional questions, please contact the Division of Molecular Biology Laboratory at (410)767-5819.