



STATE OF MARYLAND

DHMH

Maryland Department of Health and Mental Hygiene  
201 W. Preston Street • Baltimore, Maryland 21201

Martin O'Malley, Governor – Anthony G. Brown, Lt. Governor – Joshua M. Sharfstein, M.D. Secretary

Laboratories Administration  
Robert A. Myers, Ph.D., Director

**To:** CT/GC NAAT Approved Submitters  
**From:** Susan Taylor, PHLS Lead, Chlamydia Laboratory  
**Through:** Heather Peters, PHLS Supervisor, Virus Isolation/Chlamydia Laboratory  
Dr. Maria Paz Carlos, Division Chief, Virology and Immunology  
**Date:** May 23, 2012  
**Re:** Chlamydia/GC NAAT Urine Collection Protocol Reminder

*SMT 5-23-12*

*HSP 5/23/12  
MC 5/23/12*

It has been over a year since the Chlamydia Laboratory updated the urine collection procedure for the nucleic acid amplification test (NAAT) to include an "in clinic" transfer of the urine from the collection cup into a capped tube for transport to the lab. Everyone did an excellent job with this transition. At this time, it is prudent to once again stress the importance of contamination control. This is a reminder that one piece of Chlamydial DNA transferred into another sample is theoretically enough to cause the contaminated sample to test positive. Below is a list of measures that must be followed to maintain the integrity of your urine samples and prevent cross-contamination. Please take a moment to review them directly with the staff members responsible for the transfer of urine from the cup into the transport tube.

- The transfer of the urine from the cup to the tube must be performed by a trained staff member, not the patient.
- Wear clean gloves and change them whenever they get wet. Gloves are worn to protect the sample as much as to protect the wearer.
- Open only one cup of urine at a time.
- Maintain the sterility of the transfer pipette by handling it only by the bulb. If you need to set it down, slide it back into the packaging. If you drop it, or in any way compromise its sterility, discard it and open another one. Transfer pipettes must be used for a single specimen only and then discarded.
- Do not use a cap if the foil is not intact. Handle the cap only by the outside edges being careful not to contaminate the inside. Tighten the cap on the tube to prevent leakage.

**AND MOST IMPORTANTLY:**

- **Daily** clean the work area with 20% household bleach. This should be prepared freshly **each day** by diluting one part bleach in four parts water. The bleach that you dilute should start with at least a 5% concentration of sodium hypochlorite. Check the label. Dampen some disposable towels with the diluted bleach solution, wipe the surfaces, let it sit a few minutes, and then follow with water. The water step is important to physically remove any DNA. **Cleaning with a disinfectant is not a substitute for bleach.** The purpose is not as much to kill the organism as it is to destroy the DNA. Disinfectant may not accomplish that goal.

If your facility does not allow bleach, you should cover your work space with disposable towels and change them every day. Undiluted hydrogen peroxide may be used as a substitute for bleach, but a fresh bottle must be opened every eight days.

Please refer to the CT/GC NAAT urine collection and handling procedures posted on the MD DHMH Laboratories Administration website: <http://dhmh.maryland.gov/laboratories/docs/Qx%20urine%20coll.%20proc.%20clinic%20transfer%201-28-11.pdf>

Monitor your positivity rate for unusual or unexpected increases. If cross-contamination is suspected, review your technique, clean thoroughly with 20% bleach and notify the laboratory at 410-767-6154 so we can partner with you to ensure the integrity of your urine samples.

P.O. BOX 2355 • Baltimore, Maryland 21203-2355  
410-767-6100 • TTY for Disabled - Maryland Relay Service 1-800-735-2258  
Toll Free 1-877-4MD-DHMH • Web Site: [www.dhmh.state.md.us/labs/](http://www.dhmh.state.md.us/labs/)